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# Sex Determination in Plants and its Contribution to Genetic Variability

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Abstract: Sex determination is a process that leads to the physical separation of male and female gamete-producing structures to different individuals of a species. Sexual reproduction is an ancient feature of eukaryote life, yet the sexes as we currently recognize them are relative late comers in the evolution of sex. Sex determination systems in plants have evolved many times from hermaphroditic ancestors (including monoecious plants with separate male and female flowers on the same individual), and sex chromosome systems have arisen several times in flowering plant evolution. Sex chromosome evolution is intimately connected with Y chromosome degeneration. Most current understanding of how the distinctive properties of Y chromosomes evolved comes from theoretical work on the evolution of genomic regions with low recombination. The identification of sex chromosomes in plants is problematic because most of them do not differ morphologically from autosomes or from one another. For example in some species, such as Actinidia deliciosa var. deliciosa, X and Y chromosomes are too small to support observations of their distinguishing characteristics.) In the majority of plants, male and female organs are formed and developed simultaneously, but only up to a point when the growth of either set of sex organs is inhibited. Inhibition of sexual development can vary in character so that in most cases, sexual development is inhibited by the absence of cell division. In many species of bryophytes, heterothallism (unisexuality) has been correlated with the presence of sex chromosomes. Although the extent of heterothallism and sex chromosomes in the bryophytes has not been assessed systematically, this is the only known group of homosporous plants that uses sex chromosomes in sex determination. To date, studies of bryophyte sex determination have focused on the heterothallic liverwort Marchantia polymorpha. Many dioecious species, including those with well-developed sex chromosomes, show 'leaky' or 'inconstant' gender expression, with males and females producing flowers of the other gender. Far from being an aberrant feature of plant reproduction, such phenotypes are typically fully functional for both sexes. The quantitative distribution of gender in a population is under strong selection and can evolve rapidly when circumstances change for example, when males are lost from a population. In angiospermic plant, monoecious species are grouped into those that produce only unisexual male and female flowers on the same plant, and those that produce both unisexual and perfect flowers on the same plants.

Keywords: Sex, chromosome, plant.

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# INTRODUCTION

Sexual reproduction is an ancient feature of eukaryote life, yet the sexes as we currently recognize them are relative late comers in the evolution of sex (Beukeboom and Perrin, 2014). The ability to reproduce through sex (fusion of haploid gametes) evolved in the common ancestor of all eukaryotes, but did not involve separate sexes; each individual was able to exchange genetic material with any other of the same species, as is still the case in many modern-day unicellular eukaryotes (Lahr *et al.*, 2011). Sex determination is a process that leads to the physical separation of male and female gamete-producing structures to different individuals of a species. Sexual systems in plants are almost overwhelmingly diverse, and understanding their origin and evolution requires data from many different fields, including ecology, developmental biology, and genetics. Sexual system is distribution and function of gamete-producing morphological structures (Sakai AK *et al.*, 1999). Genetic control of sex determination is becoming well understood in several animal systems, particularly *Drosophila melanogaster*, *Caenorhabditis elegans* and mammals. In plants, understanding the sex determination system is closely connected with understanding how separate sexes evolved, and current theoretical ideas about this also illuminate the evolution of sex chromosomes. Angiosperms are also of particular interest for empirical studies of sex chromosome evolution, because they probably evolved separate sexes repeatedly and relatively recently. Other plants, particularly Bryophytes (Okada *et al.*, 2001), also have interesting independently evolved sex chromosomes.

In angiosperms, a sex-determining process is manifest in species that are monoecious, in which at least some flowers are unisexual but the individual is not, or dioecious, in which unisexual plants produce flowers of one sex type. In plants that produce no flowers and are homosporous, sex determination is manifest in the gametophyte generation with the production of egg- and sperm-forming gametangia on separate individual gametophytes. The determinants of sexual phenotype in plants are diverse, ranging from sex chromosomes in Marchantia polymorpha and Silene latifolia to hormonal regulation in Zea mays and Cucumis sativa to pheromonal cross-talk between individuals in Ceratopteris richardii (Geber *et al.*, 1999).

# **2. LITRETURE REVIEW**

## 2.1 Evolution of sex chromosomes

The theory outlined the evolution of a rarely recombining chromosome region containing the sex determining genes, an incipient sex chromosome system. The female haplotype carries a recessive malesterility allele, while the dominant male-determining chromosome would carry female-sterility alleles. Sex chromosome evolution is intimately connected with Y chromosome degeneration. Most current understanding of how the distinctive properties of Y chromosomes evolved comes from theoretical work on the evolution of genomic regions with low recombination. Such regions are subject to several processes, given a sufficiently high rate of deleterious mutations (Charlesworth and Charlesworth, 2000).

Westergaard (Yakubov, *et al.*, 2005) grouped plant sex chromosomes into three types to illustrate different evolutionary stages. The earliest stage of sex chromosome evolution was characterized by having a viable YY genotype where the Y differs from the X only in its sex determination genes. This condition is represented by *Ecballium* and asparagus. The second stage has a YY genotype that is inviable, but the Y chromosome plays a decisive role in sex determination; this condition is represented by papaya and white campion (dephLF, *et al.*, 2010 and Lewis D. 1942). In the third stage the Y chromosome is irrelevant to sex determination and sex is determined by the X: autosome ratio as exemplified by sorrel.

Upon the evolution of a sex-determining gene, the Y chromosome is expected to begin a process of degeneration because of its reduced population size (between one male and one female, there is only one copy of a Y chromosome and 3 copies of X chromosomes) and reduced recombination that sometimes evolves between the X and Y chromosomes; both factors reduce the effective population size of the Y chromosome and reduces efficacy of selection, which, in turn, can lead to its degeneration. Selection to reduce recombination may be a byproduct of sexual antagonism (SA), where different SA alleles provide higher fitness in males and females; for example, reduced recombination between the sex-determining region and a SA locus can lead to associations between a male benefit (female-detriment) allele and the Y chromosome, and thereby restrict the allele's benefits to males and not expose females to its costs. Cessation of recombination often occurs through inversions that isolate increasingly large stretches of the Y chromosome (Charlesworth et al., 2005). There is evidence that plant sex chromosomes derive from a pair of autosomes (Nicolas et al., 2005; Filatov 2005; Bergero and Charlesworth 2009). Theory predicts that at least two closely linked sex-determining genes are necessary for the birth of sex chromosomes; the somodel" called "two-gene (Charlesworth and Charlesworth 1978) a male sterility mutation (recessive in X/Y systems, dominant in Z/W systems) and a female sterility mutation (dominant in X/Y systems, recessive in Z/W systems.

## 2.2 Molecular basis of sex determination

Despite a large body of research investigating different species, the molecular mechanisms of sex determination in plants remain relatively unknown. It is generally believed that metabolic processes and the sexual determination pathway have genetic origins. Attempts are made to identify and estimate the number of genes which determine sex in various plant species. An identified gene is often not the primary element in the sex determination mechanism (Ainsworth 2000), but only one of the many genes in the mechanism's cascade. Frankel and Galun (1977) proposed the key gene theory to explain the sex determination mechanisms in plants. When triggered, the key gene activates a cascade of other genes whose expression leads to the development of the respective sex organs. This implies that the genes conditioning the development of male and female sex organs are present simultaneously in every plant, and those sets of loci are activated subject to the allelic architecture of the key gene. A single-gene mechanism controls sex determination in plants such as Asparagus officinalis (Gao et al., 2007), Ecballium elaterium (Ainsworth 2000), Pistacia vera (Hormaza 1994) and Carica papaya (Storey 1953).

Understanding of the evolution of plant sex chromosomes and sex determination is advanced by the

use of molecular markers since several groups are searching for. The region containing the sex determining is fully homologous between the two alternative chromosomes. Both X- and Y-linked markers are now being discovered in plants with and without heteromorphic sex chromosomes (Testolin *et al.*, 1995; Harvey *et al.*, 1997; Polley *et al.*, 1997; Zhang *et al.*, 1998; Mandolino *et al.*, 1999). Most markers are, however, anonymous, and cannot tell us which X-linked loci have homologues on the Y chromosomes and which do not.

## 2.3 Sex chromosomes

Sex chromosomes are a very peculiar part of the genome that have evolved independently in many groups of animals and plants (Bull, 1983). The identification of sex chromosomes in plants is problematic because most of them do not differ morphologically from autosomes or from one another (Spinacia oleracea, Asparagus officinalis) (Michalik 2009). In some species, such as Actinidia deliciosa var. deliciosa (Shirkot et al., 2002), X and Y chromosomes are too small to support observations of their distinguishing characteristics. Heteromorphic sex chromosomes have been discovered in Rumex acetosa (Kihara and Ono 1923), Melandrium album, Melandrium rubrum (Blackburn 1923), three species of the family Cannabidaceae, i.e. Humulus lupulus, Humulus japonicas and Cannabis sativa (Shephard et al., 1999a), as well as in Rumex nivalis (Stehlik and three Barrett 2005) and liverwort species: Sphaerocarpos donnellii, Sphaerocarpos texanus (Allen 1917) and Marchantia polymorpha (Bischler 1986). An occurrence of heteromorphic sex chromosomes is also suspected in mosses of the genus Macromitrium (Ramsay 1966).

In diploid species with a chromosomal mechanism of sex differentiation, males are generally heterogametic (XY) and females are homogametic (XX). A rare sex determination mechanism is observed in polyploid, dioecious species of the genus Fragaria where female individuals are heterogametic (Dellaporta and Calderon-Urrea 1993). Sex determination is much simpler in haploids where males and females have completely different sex chromosomes: Heteromorphic sex chromosomes are rarely found in angiosperms but have been reported in a number of plant species including Rumex, Cannabis, Humulus, and Silene (Parker, 1990). In dioecious Silene, males are the heterogametic sex (XY) and females are homogametic (XX) (Westergaard, 1948). The X chromosome appears to be essential in both males and females because only monoploid females can be obtained by in vitro techniques (Ye et al., 1991). Application of hormones, including GA, auxins, and cytokinins, does not result in sex conversion. However, the presence of a single Y chromosome can suppress female development when three X chromosomes are present. Higher X copy number overcomes the Y chromosome masculinization

effect (Westergaard, 1958). Autosome ratios have no profound effects on the sex determining factors present on the Y chromosome. This suggests that the Y chromosome is decisive in determining sex in Silene. Three different regions of the Y chromosome have been identified as having separate functions in sex determination (Westergaard, 1946). One end contains a genetic factor (or factors) that suppresses formation of the gynoecium, the opposite end contains a male fertility factor (or factors), and the middle region includes a gene or genes needed for anther initiation. Therefore, the Y chromosome of Silene contains complete linkage between female-suppressor and essential male sex genes. Asparagus is generally a dioecious plant, with sex determined by homomorphic sex chromosomes in which the males (XY) are the heterogametic sex (reviewed by Bracale et al., 1991). Genetic evidence suggests that asparagus is "male dominant" and contains male-activator-femalesuppressor genetic determinants (Marks, 1973).

Dioecious Mercurialis annua has homomorphic sex chromosomes with a small non recombining region despite fairly significant phenotypic differences between the sexes (Durand 1963; Russell & Pannell 2015). Homomorphic sex chromosomes are painstakingly difficult to identify and the hunt for sex chromosomes in poplars has produced controversial results, with some sture porting female heterogamete (ZW females and ZZ males; Paolucci et al., 2010; Yin et al., 2008), and other studies advocating male heterogamete (XY males and XX females; Gaudet et al., 2008; Geraldes et al., 2015; Pakull et al., 2009). The available female poplar genome sequence (Tuskan et al., 2006) did not clarify which system of sex determination is operating, and cytological analyses support for the presence of provided little heteromorphic sex chromosomes (Tuskan et al., 2012). Genetic mapping studies on Populus located the sexdetermining region in different positions (Gaudet et al., 2008; Yin et al., 2008; Pakull et al., 2009; Hou et al., 2015).

The primary function of sex chromosomes is to reinforce dioecy, a sexual system that may increase outcrossing in species lacking self-incompatibility (Charlesworth D. 1985. Sex chromosomes are rare in plants because of different factors. 1. Plants are sedentary, and their open architecture (bauplan) with numerous meristems allows them to grow sexual organs in varying numbers, depending on intrinsic and extrinsic factors. 2. Another reason for the rarity of plant sex Chromosomes is frequency of polyploidy in plants. Polyploidy may prevent sex chromosomes from evolving, because the frequency of homozygous recessive alleles in four or eight homologous chromosomes is much lower than in two homologous chromosomes (Wellmer F *et al.*, 2004).

#### 2.4 Mechanisms of Sex Determination in Plants

Most plant species are bisexual by nature. Angiosperms have two types of reproductive structures so that a plant may produce flowers containing both stamens and styles, alternatively, one individual can develop both male flowers (with stamina) and female flowers (with pistils) (Renner and Ricklefs 1995). In the majority of plants, male and female organs are formed and developed simultaneously, but only up to a point when the growth of either set of sex organs is inhibited. Inhibition of sexual development can vary in character so that in most cases, sexual development is inhibited by the absence of cell division, for example in Rumex acetosa (Ainsworth et al., 1995) and Melandrium album (Farbos et al., 1997), or necrosis of sexual organ cells, which is noted in Asparagus officinalis (Caporali et al., 1994) and Actinidia deliciosa (Harvey and Fraser 1988).

In many species of bryophytes, heterothallism (unisexuality) has been correlated with the presence of sex chromosomes (Smith, 1955). Although the extent of heterothallism and sex chromosomes in the bryophytes has not been assessed systematically, this is the only known group of homosporous plants that uses sex chromosomes in sex determination. To date, studies of bryophyte sex determination have focused on the heterothallic liverwort Marchantia polymorpha. In this species, the male and female thalli (vegetative gametophytes) look alike, although males and females can be distinguished easily by differences in the morphology of the sexual structure each produces. A gametophyte bears gametangia on stalked branches called antheridiophores (if male) or archegoniophores (if female) that arise from the upper surface of the thallus. Antheridiophores produce sperm-forming antheridia, and archegoniophores produce egg-forming archegonia. The sex of each haploid gametophyte is determined by cytological distinct sex chromosomes, with males having one very small Y chromosome and no X chromosome and females having one X chromosome and no Y chromosome (Lorbeer, 1934).

Tandrium and Bombyx, the X/A balance of Drosophila, occur both in the higher animals and among the dioecious seed plants. WESTERGAAD (1948, 1958) does not regard these as different systems but as two stages in the evolution of stable, multigenic mechanisms which ensure dioecism. GOLDSCHMIDT (1955) rejects WESTERGAAD's mutagenic hypothesis and interprets both systems as manifestations of a fundamental F/M balance in development. Both the X/Y and the X/A modes of sex determination occur among the dioecious species of the genus Rumex which thus provides opportunities for investigating the cytogenetic relationships of these differing mechanisms of sex determination within a single genus. An X/Y mechanism controls sex expression in the dioecious Rumex species which comprise the subgenus Acetosella (LOVE 1944). These species exist in a natural

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polyploid series: 2n = 14, 28, 42, and 56, but the male of each species is heterogametic for a single Y chromosome, combined respectively with 1,3,5, and 7 X chromosomes. The expression of maleness results from the strong epistatic effect of the Y chromosome. Females are homogametic with 2, 4, 6, or 8 X chromosomes corresponding to the level of polyploidy in the respective species. An X/A balance controls the expression of sexuality in plants of dioecious *Rumex acetosa* L. (*ONO* 1935; YAMAMOT109 38).

Plant gender is fundamentally a quantitative trait (Lloyd, D.G., and Bawa, K.S. (1984). Whereas individuals in most gonochoristic animals are either fully male or female, gender in plants is often better viewed on a quantitative scale, measured in terms of the relative allocation to each sexual function, or the proportion of genes transmitted through sperm versus eggs (Lloyd, D.G. (1980). Many dioecious species, including those with well-developed sex chromosomes, show 'leaky' or 'inconstant' gender expression, with males and females producing flowers of the other gender. Far from being an aberrant feature of plant reproduction, such phenotypes are typically fully functional for both sexes. The quantitative distribution of gender in a population is under strong selection and can evolve rapidly when circumstances change for example, when males are lost from a population (Dorken, M.E., and Pannell, J.R. (2009). Such shifts in sex allocation can occur at any level in the modular hierarchy of plant development, from the number of pollen grains produced in anthers, to the number of floral or inflorescence primordia that develop with male versus female functions (Lloyd, D.G., and Bawa, K.S. (1984). Such variation allows for multiple pathways to sex differentiation and potentially sex determination. While the distribution of quantitative gender in a population may have a strong genetic component, individuals can also modify their sex expression in response to context-dependent opportunities or costs (Lloyd, D.G., and Bawa, K.S. (1984). Plastic sex expression applies not only to the quantitative nature of sex allocation in functional hermaphrodites (Delph, L.F., and Wolf, D.E. (2005), but, more rarely, to complete sex change (Schlessman, M.A. (1988). In jack-in-the-pulpit (Arisaema species), young plants are fully male, whereas, in later seasons, they become fully female or hermaphroditic Bierzychudek, P. (1984). Such 'sexual diphase' is expected to evolve both if the marginal benefits of reproducing as male versus female change with plant size (Zhang, D.Y., and Jiang, X.H. (2002), and if there are additional benefits to sexual specialization at a particular point in time (e.g., if fitness is compromised by interference between simultaneous male and female functions) (Charnov, E.L., and Bull, J. (1977). Plasticity in sex allocation, and sexual diphasy specifically, indicate that sex may be determined not by a genetic dimorphism, but by the differential expression of genes shared by all

individuals in a population in response to physiological and environmental cues.

The distribution of dioecy and monoecy within the angiosperm phylogenetic tree strongly favors the evolutionary scenario in which unisexual flowers evolved from perfect flowers multiple times in the angiosperm lineage (Lebel-Hardenack and Grant, 1997; Charlesworth, 2002). There are a variety of sex determination mechanisms in the angiosperms. In angiospermic plant monoecious species here are grouped into those that produce only unisexual male and female flowers on the same plant, and those that produce both unisexual and perfect flowers on the same plant. Zea mays (maize) is an example of a monoecious species that produces only unisexual flowers in separate male and female inflorescences, referred to as the tassel and ear, respectively. Unisexuality in maize occurs through the selective elimination of stamens in ear florets (flowers) and by the elimination of pistils in tassel florets (reviewed by Irish, 1999). Two general classes of sex-determining mutants have been identified in maize, including those that masculinize ears and those that feminize tassels. The anther ear (an1) and dwarf (d1, d2, d3, and d5) mutants of maize are recessive and masculinize ears by preventing stamen abortion in the female florets (Wu and Cheung, 2000). Although the cloning of the sex-determining genes in maize demonstrates that GAs and possibly other steroid-like hormones play a pivotal role in stamen abortion and feminization of flowers, the spatial distribution of these molecules could have an effect on the sex determination process, as exemplified by a steep gradient in GA abundance along the maize shoot (Rood et al., 1980), which correlates well with the malesuppressing and female-promoting phenotypic effects of GA.

In male and female S. latifolia flowers, the gynoecium and androecium initiate but arrest development prematurely, leading to functionally unisexual flowers (Grant et al., 1994). The sexual phenotype of individuals is determined by sex chromosomes; males are heterogametic (XY) and females are homogametic (XX). Early cytogenetic studies of sex-determining mutants in S. latifolia led Westergaard (1946, 1958) to conclude that the Y chromosome is divided into three regions relevant to sex expression: one required for the suppression of female development and two required for the promotion of male development. None of these regions would be necessary for the development of female reproductive organs, because these functions would reside on the X or autosomal chromosomes. Additional sex-determining mutants have been generated recently by x-ray pollen of and selecting mutagenesis both hermaphrodites and asexual F1 progeny (Farbos et al., 1999; Lardon et al., 1999; Lebel-Hardenack et al., 2002).

Isolation of male-specific cDNAs from developing flower buds or reproductive organs has not yet led to discovery of sex determining genes (Matsunaga et al, 1996; Barbacar et al, 1997), probably because sex-determi (Hardenack et al, 1994; Ainsworth et al, 1995). This is not surprising, as these mutations change floral organ identities, whereas in unisexual flowers apparently normal reproductive organs merely stop developing, as predicted by the genetic model above. Both X- and Y-linked expressed loci have now been identified in S. latifolia. One approach is to directly search for sex-linked genes (Guttman and Charlesworth, 1998). This has identified the X-linked MROS-X (male reproductive organ specific) gene and its Y-linked homologue, MROS3-Y, which appears to have degenerated. MROS3- Y contains only a short region of homology to the MROS3-X sequence. This region has been evolving in a neutral manner, with a ratio of silent to replacement substitutions, Ka/Ks, of 0.974, close to unity, as expected for a sequence evolving without selective constraints (Nei, 1987).

### 3. CONCLUSION AND RECOMMENDATION

Sex determination in plants is a fundamental developmental process that is particularly important for economic reasons, because the sexual phenotypes of commercially important crops dictate how they are bred and cultivated. Although most crop plants are not considered model systems and sex determination is not a problem that can be addressed in the model angiosperm the economic value in manipulating the sexual phenotypes of crop plants. Recent studies of sex determining mechanisms have demonstrated clearly that angiosperms, including crop plants, have evolved a variety of sex-determining mechanisms that involve a number of different genetic and epigenetic factors, from sex chromosomes to plant hormones.

Production of unisexual flowers has evolved independently in many plant species, different and novel mechanisms may be operational. There is probably not one unifying mechanism that explains sex determination in plants. Currently, an important signpost on the road to the exploration of these mechanisms seem to be molecular sex-linked markers. Although these markers alone do not explain the molecular basis of sex determination in dioecious plants, but their number, structure of sequence or sequence homology between the typical, male and female sequences represent a foothold in the study of the mechanisms of sex determination.

Diversity of sex-determination mechanisms in plants reflects the important fact that separate sexes tend to have evolved independently in different lineages and much more recently than in most animal lineages. Plasticity and genetic polymorphism remains puzzling as why a chromosomal polymorphism that determines sex in the gametophytes of bryophytes has never replaced environmental sex determination in ferns or lycophytes. Mutations in many flower development genes have the potential to cause male or female sterility, thereby leading to monoecy, gynodioecy, androdioecy, and dioecy. DNA methylation on sex chromosomes is known to affect sex expression in plants. Sex reversal under different environmental conditions is likely under epigenetic control and needs to be explored. The often young sex chromosomes in angiosperms provide a unique system compare the nascent sex chromosomes with their homologous autosomes in sister species.

# REFERENCE

- Ainsworth, C. (2000). Boys and girls come out to play: the molecular biology of dioecious plants. *Annals of Botany*, 86(2), 211-221.
- Allen, C. A. (1917). A chromosome difference correlated with sex differences in *Sphaerocarpus*. Science 46: and female flower in *Asparagus officinalis*. Search for point of transition from hermaphroditic to unisexual development al pathway. Sexual Plant Reproduction, 7, 239-249.
- Bergero, R., & Charlesworth, D. (2009). The evolution of restricted recombination in sex chromosomes. *Trends Ecol Evol*, 24, 94-102.
- Beukeboom, L., Perrin, N., 2014. The Evolution of Sex Determination. Oxford: Oxford University Press.
- Bierzychudek, P. (1984). Determinants of gender in jack-in-the-pulpit: the influence of plant size and reproductive history. Oecologia, 65, 14–18.
- Bischler H. (1986): *Marchantia polymorpha* L. S. LAT. Karyotype analysis.– The Journal of the Hattori Botanical Laboratory, 60, 105-117.
- Blackburn, K. B. (1923). Sex chromosomes in plants. Nature 112: 687- 688. Botany, 86: 211-221.
- Bracale, I., Caporall, E., Galll, M.G., Longo, C., Merzlanl-Longo, O., Rossi, O., Spada, A., *Soave*, E., Falavlgna, A., Raffaldl, F., Maestrl, E., Restlvo, F.M., and Tassl, F. (1991). Sex determination and differentiation in *Aspefagus* officinslis L. Plant Sci. 80, 67-77.
- Buli, J. J. (1983). Evolution of Sex Determining Mechanisms (Menlo Park, CA: Benjamin Cummings).
- Caporali, E., Carboni, A., Galli, M. G., Rossi, G., Spada, A., & Marziani Longo, G. P. (1994). Development of male and female flower in Asparagus officinalis. Search for point of transition from hermaphroditic to unisexual developmental pathway. *Sexual Plant Reproduction*, 7(4), 239-249.
- Charlesworth, B., & Charlesworth, D. (2000). The degeneration of Y chromosomes. *Phil Trans Roy Soc Lond B*, 355, 1563–1572.
- Charlesworth B, Charlesworth D. 1978. A model for the evolution of dioecy and gynodioecy. Am Nat. 112:975–997

- Charlesworth, D. (1985). Distribution of dioecy and self-incompatibility in angiosperms. In *Evolution— Essays in Honour of John Maynard Smith*, ed. PJ Greenwood, M Slatkin, pp. 237–68. Cambridge: Cambridge Univ Press.
- Charlesworth, D., Charlesworth, B., & Marais, G. (2005). Steps in the evolution of heteromorphic sex chromosomes. *Heredity*, 95, 118–128.
- Charnov, E. L., & Bull, J. (1977). When is sex environmentally determined? *Nature*, 266, 228-230.
- Dellaporta, S. L., & Calderon-Urrea, A. (1993): Sex determination in flowering plants.– The Plant Cell 5: *deliciosa* by RAPD markers. *Scientia Horticulturae*, 94, 33-39.
- Delph, L. F., & Wolf, D. E. (2005). Evolutionary consequences of gender plasticity in genetically dimorphic breeding systems. *New Phytol*, 166, 119–128.
- DelphLF, ArntzAM, Scotti-SaintagneC, Scotti I. 2010. The genomic architecture of sexual dimorphism in the dioecious plant *Silene latifolia*. *Evolution* 64:2873–86.
- Dorken, M.E., and Pannell, J.R. (2009). Hermaphroditic sex allocation evolves when mating opportunities change. Curr. Biol. 19, 514– 517.
- Durand. B. (1963). Le complexe Mercurialis annua L. s.l.: une \_etude biosystematique. Annales des Sciences Naturelles, Botanique, 12, 579–736. Evolution, 59(4), 814- 825.
- Farbos, I., Oliveira, M., Negrutiu, I., & Mouras, A. (1997). Sex organ determination and differentiation in the dioecious plant *Melandrium album* (*Silene latifolia*): a cytological and histological analysis. Sexual PlantReproduction, 10, 155-167.
- Filatov, D. A. (2005). Substitution rates in a new Silene latifolia sex-linked gene, SlssX/Y. *Mol Biol Evol*, 22, 402–408.
- Frankel R., Galun E. (1997): Pollination machanisms, reproduction and plant breeding. Springer Berlin
- Gao W.J., Li R.L., Li Sh.F., Deng Ch.L., Li S.P. (2007): Identification of two markers linked to the sex locus
- Gaudet, M., Jorge, V., Paolucci, I., Beritognolo, I., Mugnozza, G. S., & Sabatti, M. J. T. G. (2008). Genetic linkage maps of Populus nigra L. including AFLPs, SSRs, SNPs, and sex trait. *Tree Genetics & Genomes*, 4(1), 25-36.
- Geber, M. A., Dawson, T. E., & Delph, L. F., eds. (1999). Gender and Sexual Dimorphism in Flowering Plants. (Berlin: Springer).
- Geraldes A, Hefer CA, Capron A. (2015) Recent Y chromosome divergence despite ancient origin of dioecy in poplars (Populus). Molecular Ecology, 24, 3243–3256.
- Goldschmidrt. B., 1955 Gowen, J. W., 1961 The genotypic basis of sex-expression in angiosperms.

Botan. Rev. 6: 227- The genetics of sex in Drosophila. pp. 55-93. Sex and Internal Secret.'ons, 300. 1st edition. Edited by E. ALLENW. illiams and Wilkins Co., Baltimore. Theoretical genetics. University of California Press. Berkeley. Genetic and cytologic foundations for sex. pp. 3-75. Sex and Internal Secretions, 3rd edition, Edited by W. C. YOUNG.

- Grant, S., Hunkirchen, B., and Saedler, H. (1994). Developmental differences between male and female flowers in the dioecious plant Silene latifolia. Plant J. 6, 471–480.
- Harvey CF, Gill, CP, Fraser LG, McNeilage MA (1997). Sex determination in *Actinidia*. 1. Sex-linked markers and progeny sex ratio in diploid *A. chinensis. Sex Plant Repro* 10: 149–154. Heidelberg New York.
- Hormaza J.I., Dol lo L., Pol i to V.S. (1994): Identification of RAPD marker linked to sex determination in
- Hou J, Ye N, Zhang D. (2015) Different autosomes evolved into sex chromosomes in the sister genera of Salix and Populus. Scientific Reports, 5, 9076.
- Irish, E. E. (1999). Maize sex determination. In Sex Determination in Plants, C. Ainsworth, ed (Oxford, UK: BIOS Scientific Publishers), pp. 183–188.
- Gao, W. J., Li, R. L., Li, S. F., Deng, C. L., & Li, S. P. (2007). Identification of two markers linked to the sex locus in dioecious Asparagus officinalis plants. *Russian Journal of plant physiology*, *54*(6), 816-821.
- Kihara, H., & Ono, T. (1923). The sex chromosomes of *Rumex acetosa*.– Zeitschrift für Induktive Abstammungs
- Lahr, D. J. G., Parfrey, L.W., Mitchell, E. A. D., Katz, L. A., & Lara, E. (2011). The chastity of amoebae: Re-evaluating evidence for sex in amoeboid organisms. Proceedings of the Royal Society B: Biological Sciences, 278, 2081–2090.
- Lebel-Hardenack, S., Hauser, E., Law, T., Schmidt, J., & Grant, S. (2002). Mapping of sex determination loci on the white campion (Silene latifolia) Y chromosome using amplified fragment length polymorphism. *Genetics*, 160, 717–725.
- Lewis, D. (1942). The evolution of sex in flowering plants. *Bio. Rev. Cambridge Philos. Soc.* 17, 46–67.
- Lloyd, D. G. (1980). Sexual strategies in plants. III. A quantitative method for describing the gender of plants. NZ J. Bot. 18, 103–108.
- Lloyd, D. G., & Bawa, K. S. (1984). Modification of the gender of seed plants in varying conditions. *Evol Biol*, 17, 255–338.
- Lorbeer, G. (1934). Die Zytologie der Lebermoose mit besonderer Berucksichtingung allgemeiner Chromosomenfragen. Jahrb. Wiss. Bot. 80, 567– 817.
- Love, A., 1944 Cytogenetic studies on Rumex subgenus Acetosella. Hereditas 30: 1-136.

- Mandolino G, Carboni A, Forapani S, Faeti V, Ranalli P (1999). Identification of DNA markers linked to the male sex in dioecious hemp (*Cannabis sativa* L.). *Theoret Appl Genet* 98: 86– 92.
- Marks, M. (1973). A reconsideration of the genetic mechanism for sex determination in Asparagus officinalis. Proc. Eucarpia Meeting on Asparagus, Versailles, pp. 122-128.
- Michal ik B., Klein M., Grzebelus D., Adamus A. (2009): Hodowla roślin z elementami genetyki I biotechnologii. Poznań: Państwowe Wydawnictwo Rolnicze i Leśne.
- Nicolas M. 2005. A gradual process of recombination restriction in the evolutionary history of the sex chromosomes in dioecious plants. PLoS Biol. 3:e4.
- Okada S, Sone T, Fujisawa M, Nakayama S, Takenaka M, Ishizaki K. (2001). The Y chromosome in the liverwort Marchantia polymorpha has accumulated unique repeat sequences harboring a male-specific gene. *Proc Natl Acad Sci USA* 98: 9454–9459.
- Pakull B, Groppe K, Meyer M. (2009) Genetic linkage mapping in aspen (Populus tremula L. and Populus tremuloides Michx.). Tree Genetics and Genomes, 5, 505–515.
- Paolucci I, Gaudet M, Jorge V. (2010) Genetic linkage maps of Populus alba L. and comparative mapping analysis of sex determination across Populus species. Tree Genetics and Genomes, 6, 863–875.
- Parker, J. S. (1990). Sex chromosomes and sexual differentiation in flowering plants. Chromosomes Today, 10, 187-198.
- *Pistacia vera* using bulked segregant analysis.-Theoretical and Applied Genetics 89: 9- 13.
- Polley A, Seigner E, Ganal MW (1997). Identification of sex in hop (*Humulus lupulus*) using molecular markers. *GENOME* 40: 357–361.
- Ramsay, H. P. (1966). *The Hepaticae* and *Anthocerotae* of North America. Volume I, New York: Columbia.
- Renner, S. S., & Ricklefs, R. E. (1995). Dioecy and its correlations in the flowering plants. *American Journal of Botany*, 82, 596-606.
- Russell, J. R. W., & Pannell, J. R. (2015). Sex determination in dioecious Mercurialis annua and its close diploid and polyploid relatives. *Heredity*, *114*(3), 262-271.
- Rood, S. B., Pharis, R. P., & Major, D. J. (1980). Changes of endogenous gibberellin like substances with sex reversal of the apical inflorescence of corn. *Plant Physiol*, 66, 793–796.
- Sakai AK, Weller SG. 1999. Gender and sexual dimorphism in flowering plants: a review of terminology, biogeographic patterns, ecological correlates, and phylogenetic approaches. In *Gender and Sexual Dimorphism in Flowering Plants*, ed.

MA Geber, TE Dawson, LF Delph, pp. 1–31. Berlin: Springer.

- Schlessman, M. A. (1988). Gender diphasy ("sex choice"). In Plant Reproductive Ecology: Patterns and Strategies, Lovett Doust, J. ed. (New York: Oxford University Press), pp. 139–151.
- Shephard, H., Parker, J., Darby, P., & Charles, C. A. (2004). Sex expression in hop (Humulus lupulus L. and H. japonicus Sieb. et Zucc.) floral morphology and sex chromosomes. In *Sex determination in plants* (pp. 139-150). Garland Science.
- Shirkot P., Sharma D. R., & Mohopatra, T. (2002). Molecular identification of sex in *Actinidia deliciosa var*.
- Smith, G. M. (1955). Cryptogamic Botany. Vol. II. Bryophytes and Pteridophytes. (New York: McGraw-Hill).
- Stehlik, I., Barrett S. C. H. (2005). Mechanism governing sex ratio variation in dioecious *Rumex nivalis*.
- Storey, W. B. (1953). Genetics of papaya. *Journal* of *Heredity*, 44, 70-78.
- Testolin, R., Cipriani, G., & Costa, G. (1995). Sex segregation ratio and gender expression in the genus *Actinidia*. *Sex Plant Repr*, 8, 129–132.
- Tuskan G, DiFazio SP, Faivre-Rampant P. (2012) The obscure events contributing to the evolution of an incipient sex chromosome in Populus: a retrospective working hypothesis. Tree Genetics and Genomes, 8, 559–571.
- Tuskan GA, Difazio S, Jansson S. (2006) The genome of black cottonwood, Populus trichocarpa (Torr. & Gray). Science, 313, 1596–1604. und Verebungslehre 39: 1-7. University Press 1-802.
- Wellmer F, Riechmann JL, Alves-Ferreira M, Meyerowitz EM. 2004. Genome-wide analysis of spatial gene expression in *Arabidopsis* flowers. *Plant Cell* 15:1314–26

- Westergaard, M. (1948). The relation between chromosome constitution and sex in the offspring of triploid Melandrium. Hereditas 34, 257-279.
- Westergaard, M. (1958). The mechanism of sex determination in dioecious flowering plants. Adv. Genet. 9, 217-281.
- Westergaard, M. (1946). Aberrant Y chromosomes and sex expression in Melandrium album. Hereditas 32, 419–443.
- Williams and Wilkins Co., Baltimore. 1288 B. W. SMITH KIHARA, H. Genetics of Bombyx and Drosophila: a comparison of materials, methods and results. Seiken ZihB, Rep. Kihara Inst. Biol. Res. 6: 15-29.
- Wu, H. M., & Cheung, A. Y. (2000). Programmed cell death in plant reproduction. *Plant Mol Biol*, 44, 267–281.
- Yakubov, B., Barazani, O., & Golan-Goldhirsh, A. (2005). Combination of SCAR primers and Touchdown-PCR for sex identification in Pistacia vera L. *Scientia Horticulturae*, *103*(4), 473-478.
- Ye, D., Oliveira, M., Veuskens, J., Wu, Y., Installe, P., Hinnisdaels, S., ... & Negrutiu, I. (1991). Sex determination in the dioecious Melandrium. The X/Y chromosome system allows complementary cloning strategies. *Plant Science*, 80(1-2), 93-106.
- Yin, T., DiFazio, S. P., Gunter, L. E., Zhang, X., Sewell, M. M., Woolbright, S. A., ... & Tuskan, G. A. (2008). Genome structure and emerging evidence of an incipient sex chromosome in Populus. *Genome research*, 18(3), 422-430.
- Zhang, Y. H., Stilio, V. S., Rehman, F., Avery, A., Mulcahy, D., & Kesseli, R. (1998). Y chromosome specific markers and the evolution of dioecy in the genus Silene. *Genome*, *41*(2), 141-147.
- Zhang, D. Y., & Jiang, X. H. (2002). Sizedependent resource allocation and sex allocation in herbaceous perennial plants. *Journal of Evolutionary Biology*, *15*(1), 74-83.

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