

Effects of Mycorrhizae and Aqueous Extracts of *Hyptis suaveolens* and *Cyperus rotundus* on Cotton Productivity in the Far North Region (Cameroon)

Abakar Abba Said^{1*}, Tchuenteu Tatchum Lucien², Dounia Désiré¹, Koulagna Issa Honoré¹, Ismael Ramza Haman³, Fadimatou Ahmadou⁴, Kosma Philippe⁵, Megueni Clautilde²

¹Department of Sustainable Agriculture and Disaster Management, Faculty of Science, University of Garoua, B.P. 346 - Garoua-Cameroon

²Department of Biological Sciences, Faculty of Science, University of Ngaoundere Cameroon

³Department of Biological Sciences, Faculty of Science, University of Maroua, P.O. Box 814 Maroua, Cameroon

⁴Institute for Medical Research and Medicinal Plant Studies, P.O. Box 13033 Yaounde Cameroon

⁵Higher Institute of Agriculture, Forestry, Water and Environment, University of Ebolowa Cameroon

*Corresponding author: Abakar Abba Said

| Received: 21.10.2025 | Accepted: 15.12.2025 | Published: 31.12.2025 |

Abstract: Cotton cultivation in Cameroun is characterized by low yields due to soil infertility and pest infestation. Sustainable improvement requires the use of mycorrhizae combined with aqueous extracts from *Hyptis suaveolens* and *Cyperus rotundus* rhizomes. The experimental design used is 7×2×2 split-plot with two factors and seven treatments: (control (Te-); chemical input (Te+); *H. suaveolens* (H); *C. rotundus* (C); mycorrhizae (M); two combinations: mycorrhizae-H. *suaveolens* (M+H) and mycorrhizae-C. *rotundus* (M+C); two varieties (Q302 and L457) and two localities (Kodek and Mouda). The treatments M+H, M+C, mycorrhizae (M), aqueous extracts of *H. suaveolens* and *C. rotundus* (C) improved cotton yield by 30,91; 29,66; 26,34; 9,96 and 7,01 % respectively compared to the Te- treatment. Similarly, the Te+, M+H, M+C, mycorrhizae (M), *H. suaveolens* extract (H) and *C. rotundus* extract (C) treatment improved cotton seed yield by 26,14; 26,67; 22,25; 18,04; 14,84 and 10,08 % compared to the Te- treatment. Overall, there were no significant differences between plots treated with the combination of mycorrhizae and aqueous extracts of *H. suaveolens* or *C. rotundus* could be an alternative to the use of chemical inputs in order to improve cotton productivity in the Far North Region of Cameroon while ensuring sustainable agriculture.

Keywords: Cotton, Productivity, Aqueous Extracts, Mycorrhizae, Cameroon.

INTRODUCTION

The cotton plant (*Gossypium hirsutum* L.) is a shrub belonging to the Malvaceae family, growing to a height of between 1 and 1,5 m. this plant plays an important role worldwide (Folefack, 2010), producing a fiber called cotton that is used to weave clothing (SODECOTON, 2015). Cotton cultivation also produces various derivatives that play an important role in human and livestock nutrition. The seeds obtained from ginning are used either as seed or in oil production.

In addition, the seed kernel has a protein content of 20 to 30 % (FAO, 2014). In Africa, its cultivation is an important source of foreign trade revenue and a vital

source of income for more than 10 million people (CTA, 2005). Approximately 2 million farmers produce an average of more than 2 million tons of cottonseed per year, or nearly 830. 000 tons of fiber at an average ginning yield of 41, 5 %. This volume of cotton fiber represents more than 15% of international trade, with a turnover ranging between 500 and 700 billion CFA francs (CMA/AOC, 2017).

In Cameroon, cotton cultivation covers an area of 85.000 Km² and involves around 200.000 producers, with an annual production of 250.000 tons of cottonseed (SODECOTON, 2018). More than two million people depend in one way or another on the cotton value chain (financial income, consumption of oil from cotton seeds,

Quick Response Code



Journal homepage:
<https://www.easpublisher.com/>

Copyright © 2025 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution **4.0 International License (CC BY-NC 4.0)** which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

Citation: Abakar Abba Said, Tchuenteu Tatchum Lucien, Dounia Désiré, Koulagna Issa Honoré, Ismael Ramza Haman, Fadimatou Ahmadou, Kosma Philippe, Megueni Clautilde (2025). Effects of Mycorrhizae and Aqueous Extracts of *Hyptis suaveolens* and *Cyperus rotundus* on Cotton Productivity in the Far North Region (Cameroon). *Cross Current Int J Agri Vet Sci*, 7(6), 163-173.

textile clothing, and livestock feed from oilseed meal) (EIB, 2017). Despite its multiple uses, cotton cultivation unfortunately faces several challenges in Cameroon, including rising input costs (chemical fertilizers, pesticides, etc.), declining soil fertility, and increasing pest infestations (Yao, 2011; CMA/AOC, 2017). In the absence of phytosanitary protection, cotton pests can cause a loss of more than 45.95% of the crop (SODECOTON, 2018). Cotton producers generally use chemical pesticides to control pests and synthetic chemical fertilizers to improve soil fertility and increase productivity. However, chemical control poses serious environmental risks and can lead to pests developing new resistances (Ouorou *et al.*, 2012). In fact, cotton cultivation accounts for only 2.4% of cultivated land but consume 16% of the world's insecticides, posing a great danger to human health and ecosystems (Helvetas *et al.*, 2008). The excessive use of chemical fertilizers and pesticides results in soil degradation, lower yields, pesticide resistance in pests, collateral damage to beneficial insects, water pollution, and a significant reduction in cotton producer's incomes (FAO, 2014). Due to their harmful effects, alternatives must be found (Renou *et al.*, 2016).

It is therefore necessary to consider management methods in local farming communities that enable the rational and sustainable exploitation of bioresources (Manlay, 2000) and increase agricultural production while protecting natural ecosystems (Megueni *et al.*, 2011). The use of mycorrhizal biofertilisers, aqueous extracts from *Hyptis suaveolens* leaves and *Cyperus rotundus* rhizomes has proven to be a solution for restoring soil fertility on the one hand reducing pest damage on the other. Mycorrhizal inoculum for cotton cultivation may help improve the plant's hydromineral nutrition and resistance to pests. Aqueous extracts from *H. suaveolens* leaves and *C. rotundus* rhizomes could be used as phytopesticides. These aqueous extracts offer a real advantage due to their low

persistence, low toxicity to humans and their mode of action (repellent or anti-appetitive) on pests (Ngamo *et al.*, 2007). However, no work has been done on the use of mycorrhizae and aqueous extracts from these plants in association with cotton cultivation. This study aims to improve cotton productivity while limiting the use of chemical inputs. Specifically, it will involve: (1) evaluating cotton germination rates; (2) evaluating cotton yields.

MATERIALS AND METHODS

Our research was conducted in the Far North region of Cameroon. Two sites were selected for fieldwork in this region. The first site is located in Kodek, about 15 km from Maroua. Its soil is sandy loam (representing 87% of the soil texture) and small part of its texture is clay (Seignobos *et al.*, 2000). The geographical coordinates of this site are: 10° 38'43" north latitude and 14° 24'27" east longitude, at an altitude of 523 m. These sites were chosen for their cotton farming practices by local farmers. The rainfall pattern is mainly monomodal, with a tendency for precipitation to decrease and temperatures to rise over the years (ONACC, 2018). The climate in the area is Sahelian, with a long dry season lasting nine months (October to June) and a short rainy season lasting three months (July to September). The average annual temperature is 39°C, with daily highs often reaching 45°C in April and May. The average annual rainfall is around 900-1000 mm (CTFC, 2011).

Cotton Varieties Tested

Two cotton varieties, IRMA Q302 and IRMA L457, of the genus *Gossypium hirsutum* (Malvaceae) were used as plant material. These two varieties were supplied by the Agricultural Research Institute for Development (IRAD) in Far North Cameroon. They were chosen because of their intensive use by farmers in the region (figure 1).

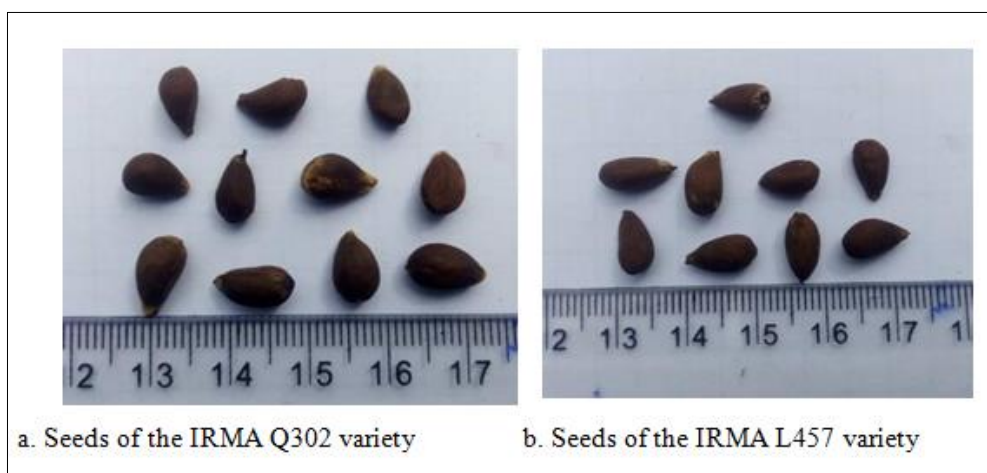


Figure 1: Cotton seeds (Abakar *et al.*, 2019)

Aqueous Extracts of *Hyptis Suaveolens* and *Cyperus Rotundus*

Aqueous extracts from the rhizome of *Cyperus rotundus* (figure 2) and the leaves of *Hyptis suaveolens* (figure 3) were used as bioinsecticides in this study. To obtain these extracts, the leaves of *H. suaveolens* and the rhizomes of *C. rotundus* were harvested and dried in the

shade for 24 hours, after which these plant parts were stored in the laboratory for 12 days before being finely ground using an electric grinder (Victoria Grain Mill-High Hopper, Medellin, Colombia). The method used to obtain the aqueous powder extracts is described by Kumar (2003) and Kosma (2011).



Figure 2: Plant, rhizomes, and aqueous extract of *Cyperus rotundus* rhizomes (Abakar *et al.*, 2020)



Figure 3: Plant, leaves, and aqueous extract of *Hyptis suaveolens* leaves (Abakar *et al.*, 2020)

The mycorrhizal inoculum used for this work comes from the collection of the Soil Laboratory of the Biotechnology centre at the University of Yaounde I. it consist of mixture of soil (clay, sand grains) and root fragments from plants containing spores of fungi of the genus *Glomus* and *Gigaspora* at a concentration of 20

spores/g of substrate (figure 4) according to jansa *et al.*, (2002), arbuscular mycorrhizal fungi of the genera *Glomus* spp. and *Gigaspora* spp. Are the best known. *Glomus* is an obligate symbiotic fungus that is not very host-specific (Bouamri *et al.*, 2006; Wang *et al.*, 2008).



Figure 4: Mycorrhizal inoculum (Abakar *et al.*, 2019)

The chemical inputs used in this experiment were purchased from the Cotton Development Corporation (SODECOTON) in Maroua. This was a single application of NPKBS complete fertilizer with the formula 21.08.12.3. This type of fertilizer is intended for cotton cultivation and is commonly used by cotton producers in the Far North (figure 5). This chemical fertilizer was applied fourteen days after sowing. The

chemical insecticide used throughout the experiment is commonly used by farmers. Its trade name is trofort 400 EC and its active ingredient is organophosphate combined with pyrethroid.

Ingredient is organophosphate combined with pyrethroid.



Figure 5: Chemical inputs (Abakar *et al.*, 2019)

The experimental design used in each location (Kodek and Mouda) is a two-factor split-plot; the main factor consists of cotton varieties (Q302 and L457 and the secondary factor consist of inputs applied to cotton (chemical fertilizer and chemical insecticide (positive control) Te+); plants that received no treatment (negative control; Te-); aqueous extract of *Hyptis suaveolens* leaves (H); aqueous extract of *Cyperus rotundus* rhizomes (C); combination of mycorrhizae and aqueous extracts of *Hyptis suaveolens* leaves (M+H) or *Cyperus rotundus* rhizomes (M+C). Each experimental unit comprises three rows of seedlings, and each row comprises ten pots, giving a total of 30 pots per elementary plot.

The sowing phase took place on June 3 and 5, 2019, in Kodek and Mouda, respectively. The sowing method used was direct seeding. Two to three cotton seeds were sown per hole at a depth of approximately 2 to 3 cm (SODECOTON, 2015). Twenty grams of mycorrhizae were added per hole (Abakar *et al.*, 2019). Mycorrhizal inoculation was carried out at sowing, and the inoculum was placed in direct contact with the seeds and throughout the cavity of the hole. To ensure that the plants were well anchored in the soil, NPKSB chemical fertilizer was applied fourteen days after sowing. This chemical fertilizer was applied directly at the base of the seedling (Deschênes, 2010). The field was maintained every two weeks by weeding, thus promoting good nutrition and water absorption.

Insecticide treatments (Trofort and aqueous solution of *Hyptis suaveolens* and *Cyperus rotundus*) were carried out on July 24, 2019, i.e., 45 days after sowing. The Trofort insecticide was applied to all aerial

parts of the plants. Treatment with aqueous extracts of *H. suaveolens* and *C. rotundus* was carried out every 7 days, for a total of 12 treatments. A Micron rotary disc sprayer was used to apply the insecticides in the field.

Determination of the emergence rate and number of nodes per cotton plant: The emergence rate of the plants was assessed 14 days after sowing. To assess the emergence rate of the plants, the number of plants that emerged was counted in order to evaluate the emergence rate per treatment. The emergence rate was calculated using the method employed by Djossin *et al.*, (2013): $ER = NEP \times 100 / TES$, where ER = emergence rate, NEP = number of plants that emerged, and TES = total number of seeds sown. The number of nodes on the main stem of the cotton plant was determined by simple counting 80 days after sowing, i.e., at capsule maturity.

$$TL = NPL \times 100 / NTP$$

TL: emergence rate;

NPL: number of plants emerged;

NTP: total number of seeds sown.

The number of nodes on the main stem of the cotton plant is determined by simple counting 80 days after sowing, i.e., when the bolls are mature.

Data Analysis:

The data were statistically analyzed using Stratigraphic Plus version 5.0 software, which performs analysis of variance (ANOVA). Duncan's test was used to assess the difference between the treatment means.

RESULTS AND DISCUSSION

Cotton Emergence Rate

The results for cotton emergence rate are shown in figures 6 and 7. Analysis of variance indicates that there is a significant difference ($P < 0.05$) between treatments in terms of cotton emergence rate. It varies between 48.66 ± 2.43 % for the treatment that received no fertiliser (Te-) of the Irma Q302 variety at the Mouda experimental site and 85.33 ± 4.21 % for the treatment enriched with mycorrhizae of the Irma Q302 variety at the Kodek experimental site. The subplots enriched with mycorrhizae had a high germination rate compared to the plots that received no treatment (Te-). This rate varies between 70.66 ± 3.51 for the Irma Q302 variety in Mouda and 85.33 ± 4.21 for the L457 variety in Kodek. However, there are no significant difference ($p < 0.05$) between varieties and study sites in terms of cotton plant emergence rates.

The percentage germination rate obtained differs from that of Djossin (2013), who obtained values between 34.75 and 56.25 % for the cotton germination rate in his study on the response of cotton (*Gossypium hirsutum* L.) to different doses of urea and sowing densities after improved fallow based on *Mucuna pruriens* at the Barre site in the commune of Zogbodomey (Abomey). This difference in results could be explained by the different agroecological zones and varieties. On the other hand, the difference in emergence rates between treatments may be due to the protection of cotton seed varieties by mycorrhizae against pathogens, mainly fungi and nematodes that attack plant seeds and roots (Fortin *et al.*, 2008; Doidy *et al.*, 2012). Indeed, mycorrhization induces an active state of immunisation in the plant, enabling it to respond more effectively to pathogen attacks (Ismail *et al.*, 2012; Jung *et al.*, 2012). The mycorrhizae added to the soil promoted the emergence of plants of the two cotton varieties used at the different trial sites.

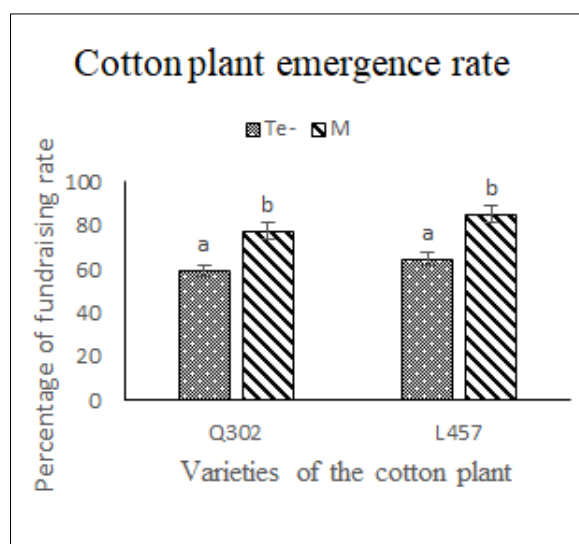


Figure 6: Emergence rate of different cotton varieties at the Kodek site

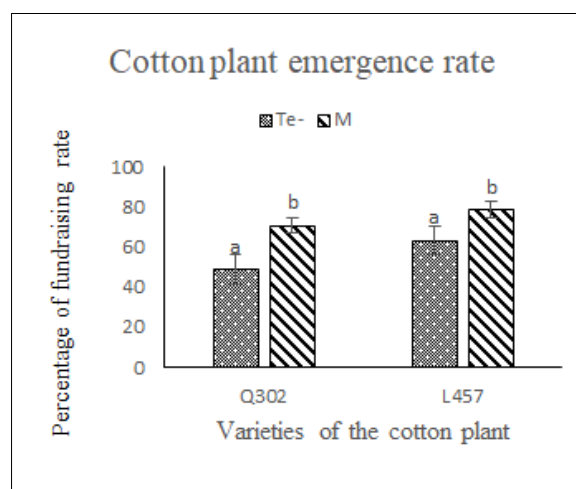


Figure 7: Emergence rate of different cotton varieties at the Mouda site

TE: Plot that received no treatment, **M:** plot enriched with mycorrhizae. The values of bands followed by the same letter in a locality are not significantly different at the 5% threshold.

Number of Nodes on the Main Stem

Figures 8 and 9 show the number of nodes on the main stem according to treatment. Analysis of variance indicates that there is a significant difference ($p < 0.05$) between treatments. The results show that the lowest number of nodes on the main stem of the cotton plant was found on plants from unfertilized plots (Te-) with an average of 12.70 ± 3.16 nodes for the Te-treatment of the L457 variety at the Mouda trial site. On the other hand, the highest number of nodes on the main stem was found on plants from the plot enriched with chemical inputs (Te+) of the Irma Q302 variety in the Kodek experimental plot. However, there were no significant difference ($p < 0.05$) between plots enriched with chemical inputs and those enriched with mycorrhizae. Overall, plots enriched with mycorrhizae increased the number of nodes on the main stem of cotton plants by 14.62 % and 15.87 % compared to untreated plots in Kodek and Mouda, respectively.

This study shows that the mycorrhizae used favorably increased the number of main stem nodes in

both cotton varieties tested, similar to plots treated with chemical inputs. Both the mycorrhizae and the chemical fertilizer applied to the soil had a positive effect on increasing the number of main stem nodes in both cotton varieties tested. However, the number of main stem nodes did not show clear differences between the study sites or between the cotton varieties. The data obtained regarding the number of main stem nodes are identical to those obtained by Sekloka (2006) and Dagbenonbakin *et al.*, (2012), who respectively obtained an average of 12 and 23 main stem nodes in their cotton studies.

This correlation is attributed to similar growing conditions and agro-ecological zones. The number of nodes on the main stem indicates healthy plant development. A higher number of nodes means the plant produces more fruiting branches. These fruiting branches bear the cotton bolls; therefore, an increase in fruiting branches would lead to an increase in cotton bolls, thus improving cotton and seed cotton yields.

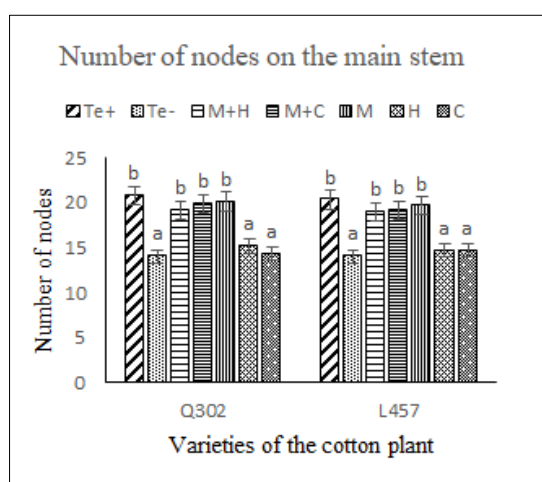


Figure 8: Number of nodes on the main stem of the cotton plant in the locality of Kodek

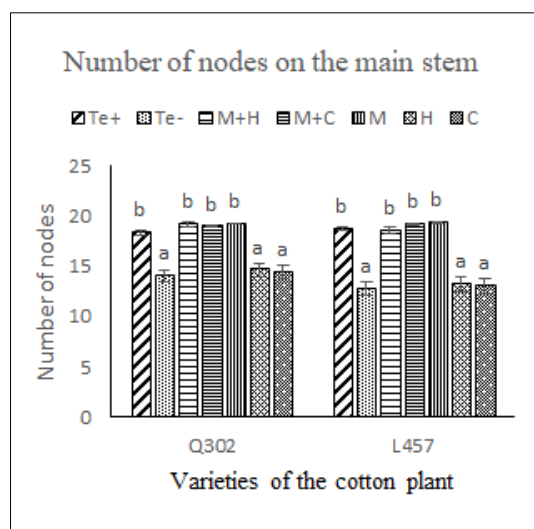


Figure 9: Number of nodes on the main stem of the cotton plant in the locality of Mouda

Cotton Fiber and Seed Cotton Yield

Effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on cotton yield.

The results of the effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on cotton fiber yield revealed a significant difference ($p < 0.05$) between the treatments (Figures 10 and 11). The plot that received no fertilizer (Te-) of the Q302 variety at the Kodek experimental site had the lowest cotton yield, with an average of 569.37 ± 12.10 kg/ha. On the other hand, the highest yield was that of the treatment of the combination mycorrhizae aqueous extracts of *H. suaveolens* (M+H) of the variety L457 in the study site of Mouda with an average of 1286.62 ± 78.45 Kg/ha. However, there were no significant differences ($p < 0.05$) between the plots treated with the mycorrhizal-aqueous extracts of *H. suaveolens* and mycorrhizal-aqueous extracts of *C. rotundus* combinations (M+C) compared to the plot treated with chemical inputs (Te+). This result shows that the M+H, M+C, mycorrhizal (M), aqueous extracts of *H. suaveolens* (H), and *C. rotundus* (C) treatments improved yield by 30.91%, 29.66%, 26.34%, 9.96%, and 7.01%, respectively, compared to the Te- treatment.

The results obtained for cotton fiber yield show that the different inputs used reduced the impact of pests

on cotton cultivation. This resulted in an increase in cotton yield compared to the Te- treatment. This result corroborates the work of Barbosa *et al.*, (2011) and Fall *et al.*, (2017), who respectively studied the toxicity of *Cyperus rotundus* extracts on *Diabrotica speciosa* and the insecticidal activity of *Callistemon viminalis*, *Melaleuca leucadendron*, and *Hyptis suaveolens* essential oils against *Sitophilus* spp., maize pests. In this regard, they demonstrated that *H. suaveolens* and *C. rotundus* extracts significantly reduce the impact of pests on crops. Thus, this improvement in cotton fiber yield would be due to the effects of extracts from *H. suaveolens* and *C. rotundus* which reduced the impact of pests on the two cotton varieties used. Similarly, mycorrhizae have not only boosted yield but, more importantly, strengthened the plant's natural defense system (Hamza, 2014; Malard, 2016; Lenoir, 2017; and Tobolbai *et al.*, 2018). Furthermore, genotypic characteristics should not be overlooked; the L457 variety appears to have genotypic characteristics that are more resistant to pests compared to the Q302 variety. Cotton fiber yield allows us to assess the effects of mycorrhizae and aqueous extracts of *H. suaveolens* and *C. rotundus* on cotton yield. In this study, the combination of mycorrhizae and *H. suaveolens* extracts proved beneficial to cotton fiber yield in the study sites.

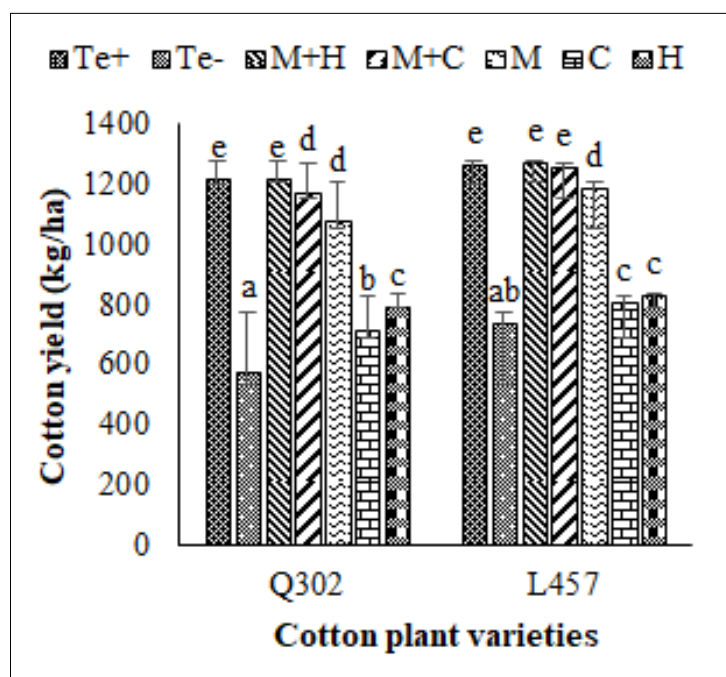


Figure 10: Effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on cotton fiber yield in the Kodek locality

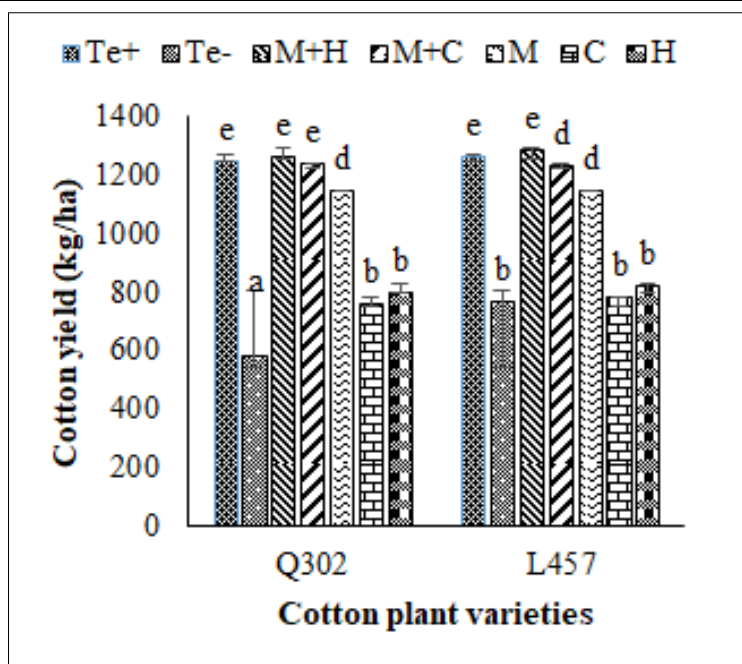


Figure 11: Effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on cotton fiber yield in the Mouda locality

Te+: plot treated with chemical fertilizer and chemical insecticide; Te-: plot without inputs; H: plot treated with aqueous extract of *H. suaveolens*; C: plot treated with aqueous extract of *C. rotundus*; M+C: plot combined with mycorrhizae and aqueous extract of *C. rotundus*; M+H: plot combined with mycorrhizae and aqueous extract of *H. suaveolens*; M: plot treated with mycorrhizae.

Effects of Mycorrhizae and Aqueous Extracts of *Hyptis Suaveolens* and *Cyperus Rotundus* on Seed Cotton Yield

Statistical analysis of the data on the effect of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on seed cotton yield reveals significant differences ($p < 0.05$) between the treatments (Figures 12 and 13). The lowest seed cotton yield was obtained from the unfertilized plot (Te-) of the Q302 variety in the Kodek locality, with an average of 1012.38 ± 68.13 kg/ha. However, the highest yield came from the plot treated with the combination of mycorrhizae-aqueous extracts of *H. suaveolens* (M+H) of the L457 variety in the Kodek site with an average of 1826.35 ± 51.27 Kg/ha. However, there are no significant differences between the plots treated with the combination of mycorrhizae-extracts of *H. suaveolens* and that of mycorrhizae-aqueous extracts of *C. rotundus* (M+C) compared to the plot treated with chemical inputs (Te+).

Overall, treatments Te+, M+H, M+C, M, H, and C improved seed cotton yield by 26.14%, 26.67%, 22.25%, 18.04%, 14.84%, and 10.08%, respectively, compared to treatment Te-. This result demonstrates that mycorrhizae and extracts of *H. suaveolens* and *C.*

rotundus improved cotton productivity. This corroborates the work of several authors who have found that extracts of *H. suaveolens* and *C. rotundus* reduce pest damage to crops (Fall *et al.*, 2017; Revi *et al.*, 2017; Sane *et al.*, 2018; Vijender *et al.*, 2018; and Abakar *et al.*, 2020). This protection is thought to be due to secondary metabolites, particularly alkaloids, flavonoids, terpenoids, and phenolic compounds present in these extracts of *H. suaveolens* and *C. rotundus*. Similarly, mycorrhizae introduced into the soil appear to have enhanced the plant's virulence against *H. armigera* (Hamza, 2014; Abakar *et al.*, 2020). Furthermore, mycorrhization induces an active state of immunity in the plant, enabling it to be more effective in its responses to pest attacks (Ismail *et al.*, 2012; Jung *et al.*, 2012; Jochems-Tanguay, 2014). Seed cotton provides vegetable oil for human consumption and protein-rich oilseed cake, which serves as a staple food for livestock. In this study, the plot enriched with mycorrhizae and treated with extracts of *H. suaveolens* yielded a higher seed cotton yield. The mycorrhizae improved the protection of the cotton bolls against this potential pest. This protection is due to the increased concentration of secondary metabolites in the cotton bolls caused by the mycorrhizae (Abakar *et al.*, 2020).

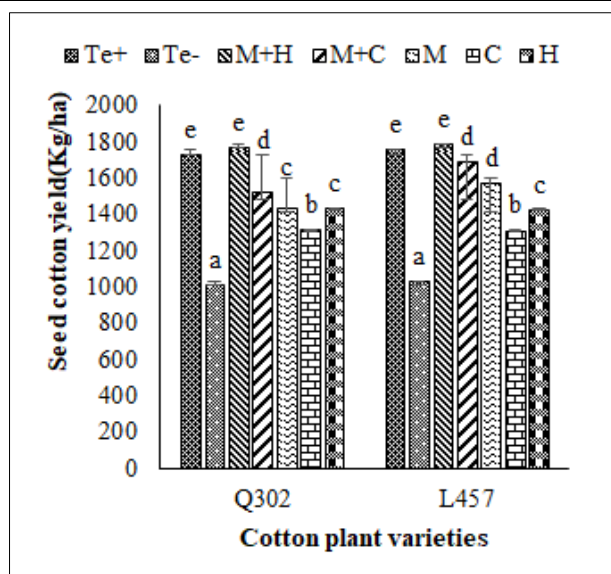


Figure 12: Effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on seed cotton yield in the Kodek locality

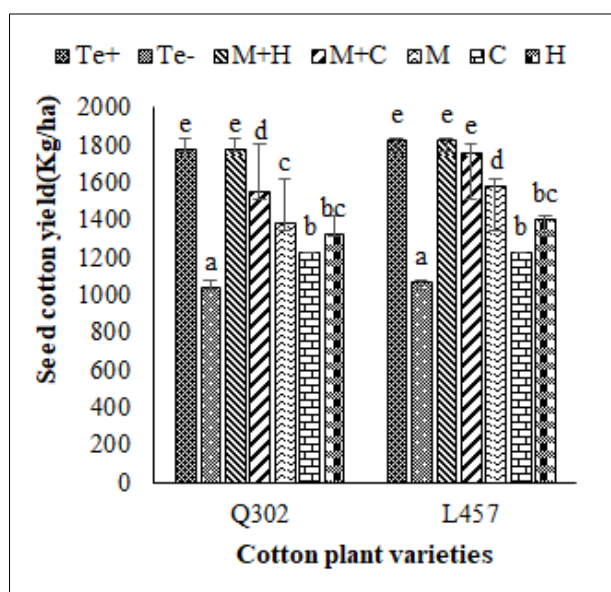


Figure 13: Effects of mycorrhizae and aqueous extracts of *Hyptis suaveolens* and *Cyperus rotundus* on seed cotton yield in the Kodek locality

CONCLUSION

Cotton yields were higher in plots treated with a combination of mycorrhizae and aqueous extracts of *Hyptis suaveolens* leaves and *Cyperus rotundus* rhizomes (M+H and M+C) compared to treatments with Te-, H, C, and M. Plots treated with the mycorrhizae and aqueous extract combination improved crop yields while reducing pest impact. The mycorrhizae facilitated the plant's water and mineral uptake, thus improving its productivity.

Similarly, the aqueous extracts used in this study limited the impact of pests on cotton cultivation. M+H and M+C treatments could be a solution for reducing the use of chemical inputs in cotton farming. Inoculating mycorrhizae at sowing and treating with

aqueous extracts of *Hyptis suaveolens* leaves or *Cyperus rotundus* rhizomes from cotton flowering (45 days after sowing) at regular 7-day intervals until boll maturation could be an alternative to the use of chemical inputs.

REFERENCES BIBLIOGRAPHIQUES

- Abakar A. S., Megueni C., Tchuenteu T. L., Kosma P., 2019. Response of two cotton varieties on mycorrhizal inoculation at Soudano- Sahelian savannah of Cameroon. *East African Scholars Journal of Agriculture and Life Sciences* pp-145-154.
- Abakar A. S., Tchuenteu T. L., Kosma P., Koulagna I. H., Kaka A. T. et Megueni C., 2020. Interaction mycorrhizes et extraits de *Hyptis suaveolens*/*Cyperus rotundus* : Bioinsecticides

- appropriés pour la protection du cotonnier contre *Helicoverpa armigera*. *Journal of Applied Biosciences* 149: 15270 - 15279
- Barbosa, Flávia S., Leite, Germano L. D., Paulino, Marney A. de Oliveira, Guilherme, D. de Oliveira, Maia, Janini T. L. S., & Fernandes, Rodrigo C. (2011). Toxicity of extracts of *Cyperus rotundus* on *Diabrotica speciosa*. *Acta Scientiarum. Agronomy*, 33(4), 607-611
 - BEI, 2017. La Banque européenne d'investissement, la Délégation de l'Union européenne et la Cellule d'appui à l'Ordonnateur national du Fonds européen de développement discutent du développement du secteur cotonnier du Cameroun. Caon Pp : 1-2
 - Bouamri R., Dalpé Y., Serrhini M. et Bennani A., 2006. Arbuscular mycorrhizal fungi species associated with rhizosphere of *Phoenix dactylifera* L. *African Journal of Biotechnology*, 16: 510-516.
 - Centre Technique de la Forêt Communale (CTFC), 2011. Etat des lieux sur les réserves forestières dans la Région de l'Extrême-Nord
 - CMA/AOC, 2017. Quel avenir pour le coton en Afrique de l'Ouest et du Centre. 33 p.
 - Dagbenonbakin G. Djondang K., Havard M. 2012. Impacts des changements de politique dans un contexte de crise mondialisée sur les acteurs des filières cotonnières d'Afrique Centrale. *Cah Agric* 19 (1) : 21 - 27
 - Deschênes M-L., 2010. Augmentation du potentiel de croissance d'une plantation d'épinette blanche. *Rapport Technique-Projet N°093, Volet II 2009-2010 Québec/ Canada*. 240p.
 - Djossin A., 2013. Réponse du cotonnier (*Gossypium hirsutum* L.) A différentes doses d'urée et densités de semis après une jachère de *Mucuna pruriens* L. Sur terre de barre dans la commune de zogbodomey. Rapport de fin de formation pour l'obtention de la licence professionnelle en sciences agricoles. Université d'Abomey-Calavi Pp : 1-73
 - Doidy J, Grace E, Kühn C, Simon-Plas F, Casieri L, Wipf D. 2012. Sugar transporters in plants and in their interactions with fungi. *Trends in plant science* 17: 413–422.
 - Fall R, Ngom S, Sembène ASM, 2017. Activité insecticide par fumigation des huiles essentielles de *Callistemon viminalis*, *Melaleuca leucadendron* et *Hyptis suaveolens* contre *Sitophilus* spp. ravageurs du maïs. Pp :31-36.
 - FAO (2014). «Analyse des incitations par les prix du coton au Mali 2005-2012». Note de synthèse FAO, 49 p.
 - FAO, 2014. Gestion intégrée de la production et des déprédateurs du coton ; Guide du facilitateur pour les Champs écoles des producteurs. Pp : 1-89
 - Folefack D. P. 2010. Coordination des acteurs dans un contexte de crise : le cas de la filière coton au Cameroun depuis 1990. Thèse de doctorat, Economie, Université Rennes 2 Haute Bretagne (France).
 - Fortin J.A., Plenchette C. et Piché Y., 2008. Les mycorhizes-la nouvelle révolution verte. *Les Éditions Multi Mondes*. Versaille. Pp : 1-131
 - Hamza N., 2014. Application des mycorhizes arbusculaires en culture maraîchère cas de la pastèque (*Citrus lanatus*), Magister en Biologie et Physiologie Végétale.
 - Helvetas, 2008. Guide de production du coton biologique et équitable. un manuel de préférence pour l'Afrique de l'Ouest. <http://www.organiccotton.org> Pp : 1-49.
 - Ismail, Y. and Hijri, M. (2012). Arbuscular mycorrhisation with *Glomus irregulare* induces expression of potato PR homologues genes in response to infection by *Fusarium sambucinum*. *Functional Plant Biology*, 39 (3), p. 236-245
 - Jansa J., Mozafar A., Anken T., Ruh R., Sanders I. R. et Frossard E., 2002. Diversity and structure of AMF communities as affected by tillage in a temperate soil. *Mycorrhiza* 45 : 225–234.
 - Jochems-Tanguay L., 2014. Les inoculants mycorrhiziens pour une agriculture québécoise plus productive et moins dépendante aux engrais minéraux phosphatés. Essai présenté au Centre universitaire de formation en environnement et développement durable. Pp : 1-99
 - Jung, S. C., Martinez-Medina, A., Lopez-Raez, J. A. and Pozo, M. J. (2012). Mycorrhiza-Induced Resistance and Priming of Plant Defenses. *Journal of Chemical Ecology*, 38 (7), p. 651-664
 - Kosma P, Ambang Z, Boyogueno ADB, Ten Hoopen M, Kuate J, Akoa A (2011). Assessment of nematicidal properties and phytochemical screening of neem seed formulations using *Radopholus similis*, parasitic nematode of plantain in Cameroun. *Crop Protection* 30: 733-738
 - Kumar H. 2003. Association des Producteurs de Coton Africains Professionnalisation des organisations et défense des intérêts des producteurs de coton. *Global, Revue Africaine sur le Commerce et le Développement* 4 : 13 - 15.
 - Kumar, V. (2016). Weeds in Tropics: Problems and prospects. *Van Sang yan* 3(2): 1-6.
 - Lawson, A.J. 2008. Effet de différentes pratiques de taille sur l'amélioration des performances agronomiques du cotonnier *Gossypium hirsutum*.
 - Lenoir I., 2017. Intérêt de la symbiose mycorrhizienne à arbuscules dans la phytoremédiation des sols historiquement contaminés par les hydrocarbures : de la protection à la dissipation. *Biotechnologies*. Université du Littoral Côte d'Opale, 2015. Français. Pp : 1-231,
 - Malard A., 2016. Les mycorhizes, les plantes, la biodiversité et la Permaculture. Pp : 1-9
 - Manlay R., 2000. Dynamique de la matière organique à l'échelle d'un terroir agro-pastoral de la savane ouest-africaine (Sud-Sénégal). *Thèse Doctorat ès sciences de l'environnement*. Ecole

- Nationale du Génie Rural, des Eaux et Forêts. Université de Montpellier, 246 p.
- Megueni C., Awono E. T. et Ndjouenkeu R., 2011. Effet simultané de la dilution et de la combinaison du *Rhizobium* et des mycorhizes sur la production foliaire et les propriétés physico-chimiques des jeunes feuilles de *Vigna unguiculata* (L.) Walp. *Journal of Applied Biosciences* 40: 2668-2676.
 - Ngamo L.S.T et Th. Hance., 2007. Diversité des ravageurs des denrées et méthodes alternatives de lutte en milieu tropical. Pp 215-218.
 - ONACC, 2018. Gestion des risques en agriculture : évaluation et conception des politiques. Editions OCDE, 287 p
 - Ouorou K. D. K., Denis, Assouan D., Isabelle A. G., Manuele T., 2012. Réponse des stades larvaires de *Helicoverpa armigera* (Hübner) (Lepidoptera : Noctuidae) à l'application de champignons entomopathogènes *Metarhizium anisopliae* et *Beauveria bassiana*. Pp : 283-293
 - Renou A. & Brévault T., 2016. Ravageurs et maladies du cotonnier, et gestion intégrée des ravageurs. In: Crétenet M. & Gourlot J.-P., eds. Le cotonnier. Versailles, France : Quæ ; CTA ; Gembloux, Belgique : Les Presses agronomiques de Gembloux, 109-154
 - Revi Yenti, Farida Rahim, Dan Agustin Nofrianti, 2017. Penetration Test of Gel ethanol extract of *Cyperus rotundus* L. Pp 58-69
 - Sané B., Badiane D., Guèye M.T., Faye O. 2018. Évaluation de l'efficacité biologique d'extraits de neem (*Azadirachta indica*) comme alternatif aux pyrèthroïdes pour le contrôle des principaux ravageurs du cotonnier (*Gossypium hirsutum*) au Sénégal. *International Journal of Biological and Chemical Sciences*, 12 (1): 157-167.
 - Seignobos C. et Olivier Iyébi-Mandjek, 2000. Atlas de la province Extrême-Nord Cameroun. Pp : 1-172 ;
 - Sekloka E. 2006. Amélioration de l'efficacité de la sélection pour le rendement en coton graine du cotonnier *Gossypium hirsutum* L. dans un contexte de nouveau itinéraire technique. Thèse Ecole Nationale Supérieure Agronomique de Rennes. Pp.192.
 - SODECOTON, 2018. Rapports annuels, rapports du service suivi-évaluation, rapports du projet Eau-Sol-Arbre, Pp : 1-45
 - SODECOTON/DPA N°001/2015. Fiche technique de la culture cotonnière. Pp1-13
 - Tobolbai R, Adamou S, Ngakou A, 2018, Morphological and structural diversities of Ingenous endomycorhiza communities associated to maize (*Zea mays* in Northern Camerounian soils. Pp. 18
 - Vijinder S., Mohammed A., Archana N. end Shahnaz S., 2018. Analysis and antimicrobial activity of the essential oil of *Cyperus rotundus* L. rhizome. *Journal of Medicinal Plants Studies*. PP: 101-105 Pp.
 - Wang Y., Vestberg M., Walker C., Hurmer T., Zhang X. et Lindström K., 2008. Diversity and infectivity of arbuscular mycorrhizal fungi. *Agricultural soils of the Sichuan*. 18: 59-68.
 - Yao K., 2011. Coton Africain : Défis, Enjeux et perspectives. Rencontre panafricain sur le coton. Paris, France. Pp 5-19.